MIDDLEBROOK 7H11 AGAR

Cat. no. C36	Middlebrook 7H11 Agar, 20x125mm Tube, 10ml Slant	20 tubes/box		
Cat. no. C36BX	Middlebrook 7H11 Agar, 20x125mm Tube, 10ml Slant	100 tubes/box		
Cat. no. X25	Middlebrook 7H11 Agar, 50ml HardyFlask™, 12ml Slant	20 flasks/box		
Cat. no. W35	Middlebrook 7H11 Agar, 15x100mm Plate, 28ml	10 plates/bag		
Cat. no. C38	Middlebrook 7H11 Selective Agar, 20x125mm Tube, 10ml Slant	20 tubes/box		
Cat. no. C38BX	Middlebrook 7H11 Selective Agar, 20x125mm Tube, 10ml Slant	100 tubes/box		
Cat. no. X28	Middlebrook 7H11 Selective Agar, 50ml HardyFlask™, 12ml Slant	20 flasks/box		
Cat. no. J75	Cat. no. J75 Middlebrook 7H11 / 7H11 Selective Agar, 15x100mm Biplate, 15ml/15ml			
Cat. no. W40	Middlebrook 7H11 Selective Agar, 15x100mm Plate, 28ml	10 plates/bag		

INTENDED USE

Hardy Diagnostics Middlebrook 7H11 Agar is recommended for use in the isolation and cultivation of *Mycobacterium* species. (1,8)

SUMMARY

In 1947 Dubos and Middlebrook formulated a media (7H9) containing albumin and oleic acid which enhanced the growth of tubercle bacilli, and protected the organisms against a variety of toxic agents. ⁽⁵⁾ In 1958, Middlebrook and Cohn improved this formulation and developed a media (7H10) which allowed for more luxuriant and rapid growth of *Mycobacterium* species. ⁽⁹⁾ In 1968, Cohn incorporated casein hydrolysate into 7H10 medium to stimulate the growth of mycobacteria that would not otherwise grow on this medium. This formulation was then designated as 7H11 Agar, and is recommended over 7H10 Agar. ^(4,7)

Middlebrook 7H11 Agar contains inorganic compounds that supply essential growth stimulating inorganic salts, as well as vitamins and necessary co-factors. Glycerol is provided as a source of carbon and energy for the tubercle organisms. Sodium citrate is converted to citric acid, which holds inorganic cations in solution. Casein hydrolysate is incorporated into 7H11 Agar as a growth stimulant for strains of drug resistant *Mycobacterium tuberculosis*. ^(2,8) Malachite green is added as a selective agent, which partially inhibits the growth of other bacteria. Biotin helps stimulate the revival of damaged target organisms. It is also involved in a variety of carboxylation and decarboxylation reactions. OADC Enrichment contains the following additives required for growth: albumin to protect the tubercle bacilli against toxic agents; oleic acid, a fatty acid utilized in the metabolism of the organism; sodium chloride to maintain osmotic equilibrium; catalase to destroy any toxic peroxides in the medium; and dextrose as an energy source.

Middlebrook 7H11 Selective Agar was formulated in 1972 for use in isolating and cultivating Mycobacterium species

from specimens containing mixed flora. Using antimicrobics as selective agents, Mitchison, et al. formulated an agar medium (7H10 Base) designed to isolate mycobacteria. This 7H10 basal medium was later modified by Mitchison, et al. using 7H11 as the base. The antimicrobics in 7H11 Selective Agar are as follows: carbenicillin and polymyxin B, which are active against most of the Enterobacterales; trimethoprim lactate, which inhibits *Proteus* species; and amphotericin B which is active against yeasts.

FORMULA

Ingredients per 900ml of deionized water:*

Glycerol	5.0ml
Disodium Phosphate	1.5gm
Monopotassium Phosphate	1.5gm
Pancreatic Digest of Casein	1.0gm
L-Glutamic Acid	0.5gm
Ammonium Sulfate	0.5gm
Sodium Citrate	0.4gm
Magnesium Sulfate	50.0mg
Ferric Ammonium Citrate	40.0mg
Malachite Green	1.0mg
Pyridoxine	1.0mg
Biotin	0.5mg
Agar	15.0gm

OADC Enrichment:					
Bovine Albumin	5.0gm				
Beef Catalase	4.0mg				
Dextrose	2.0gm				
Sodium Chloride	0.85gm				
Oleic Acid	50.0mg				

Middlebrook 7H11 Selective Agar contains the following additional selective ingredients per liter:

Carbenicillin	50.0mg
Polymyxin B	25.0mg
Trimethoprim	20.0mg
Amphotericin B	10.0mg

Final pH 6.8 +/- 0.3 at 25°C.

* Adjusted and/or supplemented as required to meet performance criteria.

STORAGE AND SHELF LIFE

Storage: Upon receipt store at 2-8°C away from direct light. Media should not be used if there are any signs of deterioration (shrinking, cracking, or discoloration), contamination, or if the expiration date has passed. Product is light and temperature sensitive; protect from light, excessive heat, moisture, and freezing.

The expiration dating on the product label applies to the product in its intact packaging when stored as directed. The product may be used and tested up to the expiration date on the product label and incubated for the recommended quality control incubation times.

Refer to the document "Storage" for more information.

PRECAUTIONS

This product may contain components of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not guarantee the absence of transmissible pathogenic agents. Therefore, it is recommended that these products be treated as potentially infectious, and handle observing the usual universal blood precautions. Do not ingest, inhale, or allow to come into contact with skin.

This product is for *in vitro* diagnostic use only. It is to be used only by adequately trained and qualified laboratory personnel. Observe approved biohazard precautions and aseptic techniques. All laboratory specimens should be considered infectious and handled according to "standard precautions." Refer to the document "Guidelines for Isolation Precautions" from the Centers for Disease Control and Prevention.

For additional information regarding specific precautions for the prevention of the transmission of all infectious agents from laboratory instruments and materials, and for recommendations for the management of exposure to infectious disease, refer to CLSI document M-29: *Protection of Laboratory Workers from Occupationally Acquired Infections: Approved Guideline.*

Sterilize all biohazard waste before disposal.

Refer to the document "Precautions When Using Media" for more information.

PROCEDURE

Specimen Collection: Infectious material should be submitted directly to the laboratory without delay and protected from excessive heat and cold. Consult listed references for information on specimen collection. (1-3,6,7,13)

Method of Use:

- 1. Inoculate the Middlebrook 7H11 Agar with specimen, after decontamination and neutralization, according to test procedures recommended by the Centers for Disease Control (CDC). Consult listed references for methods. (1-3,6,7,13)
- 2. Incubate medium in a CO_2 atmosphere at 35-37°C. Protect from light. Tubed media should be incubated for one week with loosened caps to allow the circulation of CO_2 for the initiation of growth. Caps should be tightened after one week in order to prevent dehydration of media.
- 3. Examine the media within five to seven days, and weekly thereafter for up to eight weeks.
- 4. Examine plates under light for the appearance of macroscopic growth. For the rapid microcolony method, refer to the procedure outlined on the Instructions for Use (IFU) for Cat. no. SP57, Middlebrook 7H11 Agar, Thin Pour plate.
- 5. Examine tubes under light and magnifying mirror for macroscopic growth. Record and describe colony morphology on the first day growth is observed.

6. Consult appropriate references for recording the number of colonies and for aid in the biochemical identification of acid-fast bacilli. (1,2,6,13)

INTERPRETATION OF RESULTS

Consult listed references for the interpretation of growth of *Mycobacterium* species on this medium.^(1-3,6,7,13) Examine and record each type of colony morphology, pigment, and growth rate. Biochemical testing is required for definitive identification.

LIMITATIONS

It is recommended that biochemical, immunological, molecular, or mass spectrometry testing be performed on colonies from pure culture for complete identification.

Middlebrook 7H11 Agar requires incubation in a 5-10% CO₂ atmosphere in order to recover mycobacteria. Mycobacteria, for unknown reasons, are not recovered well from candle extinction jars.⁽⁷⁾

Keep inoculated media away from light or excessive heat. Exposure to such conditions results in the release of formaldehyde in the media which may inhibit or kill mycobacteria.

Refer to the document "Limitations of Procedures and Warranty" for more information.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as loops, slides, decontamination supplies, applicator sticks, pipets, incinerators, CO₂ incubator, and microscopes, etc., as well as serological and biochemical reagents, are not provided.

QUALITY CONTROL

Hardy Diagnostics tests each lot of commercially manufactured media using appropriate quality control microorganisms and quality specifications as outlined on the Certificates of Analysis (CofA). The following organisms are routinely used for testing at Hardy Diagnostics:

Toot Organisms	Inoculation	Incubation			Results	
Test Organisms	Method*	Time	Temperature	Atmosphere	Kesuits	
Mycobacterium tuberculosis H37Ra ATCC [®] 25177***	G	21 days	35°C	CO ₂ **	Growth; colonies visible in 2 weeks, mature in 3 weeks	
Mycobacterium kansasii Group I ATCC [®] 12478***	G	21 days	35°C	CO ₂ **	Growth; colonies visible in 2 weeks, mature in 3 weeks	
Mycobacterium scrofulaceum Group II ATCC® 19981***	G	21 days	35°C	CO ₂ **	Growth; colonies visible in 2 weeks, mature in 3 weeks	
Mycobacterium intracellulare Group III ATCC® 13950***	G	21 days	35°C	CO ₂ **	Growth; colonies visible in 2 weeks, mature in 3 weeks	
Mycobacterium fortuitum Group IV ATCC® 6841***	G	21 days	35°C	CO ₂ **	Growth; colonies visible in 4 days	

Mycobacterium tuberculosis ATCC® 35743	G	21 days	35°C		Growth; colonies visible in 2 weeks, mature in 3 weeks
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The above organisms are used for performance testing of Middlebrook 7H11 Agar. The following organisms are additionally tested on Middlebrook 7H11 Selective Agar:

Toot Organisms	Inoculation		Incubation		Results	
Test Organisms	Method*	Time	Temperature	Atmosphere	Results	
Escherichia coli ATCC [®] 25922	В	24hr	35°C	Aerobic	Partial to complete inhibition	
Staphylococcus aureus ATCC [®] 25923	В	24hr	35°C	Aerobic	Partial to complete inhibition	
Candida albicans ATCC [®] 10231	В	24hr	35°C	Aerobic	Partial to complete inhibition	

^{*} Refer to the document "Inoculation Procedures for Media OC" for more information.

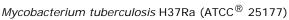
USER QUALITY CONTROL

End users of commercially prepared culture media should perform QC testing in accordance with applicable government regulatory agencies, and in compliance with accreditation requirements. Hardy Diagnostics recommends end users check for signs of contamination and deterioration and, if dictated by laboratory quality control procedures or regulation, perform quality control testing to demonstrate growth or a positive reaction and to demonstrate inhibition or a negative reaction, if applicable. Hardy Diagnostics quality control testing is documented on the certificates of analysis (CofA) available from Hardy Diagnostics Certificates of Analysis website. In addition, refer to the following document "Finished Product Quality Control Procedures," for more information on QC or see reference(s) for more specific information.

PHYSICAL APPEARANCE

Middlebrook 7H11 and 7H11 Selective Agars should appear clear, slightly opalescent, and light amber with a green hue in color.







Mycobacterium kansasii Group I (ATCC® 12478)

^{**} Atmosphere of incubation is enriched with 5-10% CO₂.

^{***} Recommended QC strains for User Quality Control according to the CLSI document M22 when applicable.



Mycobacterium scrofulaceum Group II (ATCC $^{\circledR}$ 19981) colonies growing on Middlebrook 7H11 Agar (Cat. no. C36). Incubated in CO $_2$ for 21 days at 35°C.



Mycobacterium intracellulare Group III (ATCC® 13950) colonies growing on Middlebrook 7H11 Agar (Cat. no. C36). Incubated in CO₂ for 21 days at 35°C



Mycobacterium fortuitum Group IV (ATCC $^{\otimes}$ 6841) colonies growing on Middlebrook 7H11 Agar (Cat. no. C36). Incubated in CO $_2$ for 21 days at 35°C.



Uninoculated tube of Middlebrook 7H11 Agar (Cat. no. C36).

REFERENCES

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IFU-10573[C]



1430 West McCoy Lane, Santa Maria, CA 93455, USA Phone: (805) 346-2766 ext. 5658 Fax: (805) 346-2760

Website: <u>HardyDiagnostics.com</u>

Email: TechnicalServices@HardyDiagnostics.com

Ordering Information

Distribution Centers:

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